Dietary exposure to essential oils of selected *Pinus* and *Abies* species leads to morphological changes in *Drosophila melanogaster* wings

^{(D}Vladimir J. Cvetković^{1,*}, ^{(D}Maja Lazarević¹, ^{(D}Zorica S. Mitić¹, ^{(D}Bojan Zlatković¹, ^{(D}Milica Stojković Piperac¹, ^{(D}Snežana Jevtović², ^{(D}Gordana Stojanović² and ^{(D}Vladimir Žikić¹)

¹Department of Biology and Ecology, Faculty of Sciences and Mathematics, University of Niš, Višegradska 33, 18000 Niš, Serbia ²Department of Chemistry, Faculty of Sciences and Mathematics, University of Niš, Višegradska 33, 18000 Niš, Serbia

*Corresponding author: vladimir.cvetkovic@pmf.edu.rs

Received: May 27, 2024; Revised: June 27, 2024; Accepted: July 5, 2024; Published online: July 11, 2024

Abstract: Essential oils (EOs) from conifer species are a potential source of biopesticides due to their various bioactive effects against pest insects. Since the harmful activity of pest insects often relies on their ability to fly, the disruption of wing morphology due to the use of essential oils (EOs) could be significant. This study investigated changes in wing shape and size in the model insect *Drosophila melanogaster* after exposure to EOs derived from selected conifers species. *Drosophila* larvae were treated with a diet containing 3% EOs from 6 *Pinus* and 3 *Abies* species. After completing their life cycle, the wings of adults were dissected and analyzed using the geometric morphometrics method. Changes in *Drosophila* wing morphology were observed. The potential effect of the major compounds of the tested EOs on wing morphology is discussed. We believe that this study lays the foundation for further evaluation of *Abies* and *Pinus* EOs in targeted toxicological studies against pest insects by focusing on the potential of EOs to disrupt pest activities that depend on wing morphology.

Keywords: Abies, Drosophila melanogaster, essential oils, Pinus, wing morphology

INTRODUCTION

The pine family, Pinaceae, is a significant coniferous taxon comprising 11 genera [1]. Among these is the genus Abies Mill. (firs), encompassing 52 species and the genus Pinus L. (pines) containing 133 species, representing the most diverse groups [1]. In traditional medicine of different cultures, certain essential oils (EOs) derived from Abies and Pinus have been recognized for their beneficial effects in treating respiratory infections and for muscle relaxation [2,3]. Consequently, during the last decades, there has been an increased interest in studying the chemical profiles and bioactivities of fir and pine EOs. Many studies have shown that EOs contain an array of bioactive compounds characterized by antimicrobial [4-6], molluscicidal [7], insecticidal [4-6,8,9], antiradical [10,11], antiplatelet [12], and antitumor properties [13].

The fruit fly *Drosophila melanogaster* Meigen, 1830 is an important model organism [14,15] in biological research, widely recognized as a multicellular eukaryotic insect model for numerous reasons. It is a low-cost animal model with a rapid life cycle and well-understood biology. Several assays that include *D. melanogaster* are frequently used in studies of the biological activity of plant-derived extracts [16], EOs [17-19], plant-derived oil [20], or fruit-derived compounds [21]. Furthermore, *D. melanogaster* is an effective model organism for investigating EOs as potential biopesticides for pest insect control, particularly closely related species such as *Drosophila suzukii* (Matsumura). Recent reports have highlighted the promising impact of volatile plant compounds against this invasive pest insect [22], demonstrating both insecticidal and deterrent effects [23-24].

However, recently published studies have demonstrated a significant impact of EOs extracted from conifers on the different aspects of *D. melanogaster* life cycle, such as survival rate, pupation rate, adult eclosion, and life cycle duration [4-6,25]. It was observed that adult fruit flies that survived the dietary treatment with EOs of *Abies* and *Pinus* species during the larval stage did not exhibit visible large-scale morphological changes on the body or the wings of *D. melanogaster* [4-6]. The question has been raised about whether treatments using EOs extracted from six Pinus and three Abies species affect the micro-scale morphology of D. melanogaster, particularly their wings. Based on recently published papers [26,27], one approach is to use the geometric morphometric method. Cvetković et al. [26] found micro-scale changes in wing morphology in D. melanogaster treated with TiO, nanoparticles despite their normal appearance at a larger scale. Using this sensitive method, changes in insect wings caused by active substances such as insecticides were examined in store-product coleopteran pests, including the Khapra beetle, Trogoderma granarium Everts (Coleoptera: Dermestidae) [28], Tenebrio molitor L. (Coleoptera: Tenebrionidae), Prostephanus truncatus (Horn), and Rhyzopertha dominica (F.) (Coleoptera: Bostrychidae) [29]. Furthermore, this method has demonstrated its suitability for distinguishing between groups subjected to specific treatments, as in Culicidae [30], which, like D. melanogaster, are dipterans. A recent investigation into the influence of EOs on the wing morphology of dipterans, particularly mosquitos, showed that Croton tetradenius EO influenced the wing morphometry of Aedes aegypti [27]. Changes in wing morphology can affect dipteran dispersion and flying capability [30]. Thus, it can be assumed that changes in wing morphology potentially induced by treatment with EOs could affect flying insect pests and reduce their impact on economically important crops. The aim of this study was to explore potential changes in wing shape and size in the insect model D. melanogaster following dietary exposure to EOs from selected Abies and Pinus species.

MATERIALS AND METHODS

Ethics statement

This study does not involve any live vertebrates or human subjects.

Plant material processing

Needles of six *Pinus* species (*P. halepensis* Mill., *P. heldreichii* Christ, *P. mugo* Turra, *P. nigra* J.F. Arnold, *P. peuce* Griseb. and *P. sylvestris* L.), and twigs with needles of three *Abies* species (*A. alba* Mill., *A.* × *borisiiregis* Mattf., and *A. cephalonica* Loudon) used in this

study were two years old and collected from their native populations in the Balkan Peninsula. Vernacular names for the species whose EOs were tested are P. halepensis - Aleppo pine, P. heldreichii - Bosnian pine, P. mugo - Mountain pine, P. nigra - European black pine, P. peuce - Macedonian pine, P. sylvestris - Scots pine, A. alba - Silver fir, A. x borisii-regis - King Boris fir and A. cephalonica – Greek fir. Details regarding the collection of plant material, as well as the locations and ecological conditions of the populations of studied species of Abies and Pinus have already been reported [4-6]. Voucher specimens of each taxon were deposited in the "Herbarium Moesiacum Niš" (HMN) of the Faculty of Sciences and Mathematics, Department of Biology and Ecology, University of Niš under acquisition numbers 13548, 13885, 12810, 6920, 12812, 6938, 13944, 13943, and 13999, respectively.

Isolation and chemical characterization of EOs

The isolation and determination of the chemical composition of the three *Abies* and six *Pinus* species EOs used in this study were performed and published in previous studies [4-6]. Briefly, plant material was subjected to hydrodistillation using a Clevenger apparatus. The qualitative analysis of the EOs was conducted through gas chromatography-mass spectrometry (GC-MS), while quantitative analysis involved gas chromatography-flame ionization detector (GC-FID) analysis. Both analyses were carried out under the previously presented operating parameters [31].

Bioassay: insects and their husbandry

Specimens of *D. melanogaster* were used as a model system in this experiment (Oregon stock, Bloomington Indiana University, USA). Flies in stock were reared *en masse* on a standard cornmeal-based feeding medium (agar, sucrose, yeast, water, corn meal) with the addition of methyl 4-hydroxybenzoate as a fungicide. The laboratory conditions for maintenance of the flies were standard: 25°C, 60% relative humidity, and a light regime of 12h of day and 12h of night.

Adult male and female flies used in this analysis originated from previous studies where the treatment protocol was explained in detail [4-6]. Briefly, the treatment protocol included the following steps: initially, 3-day-old larvae were obtained from young adults of the same age that were being mass-cultured. Subsequently, treatments were prepared by adding 100% pure EOs from one of the tested *Abies* or *Pinus* species into fresh cornmealbased feeding media, achieving a final concentration of 3% essential oil within the media. The 3% concentration of the EOs was selected based on prior findings, which exhibited a positive correlation with significant larvicidal effects and delayed developmental time

in *D. melanogaster* [4-6]. Therefore, it is presumed that the 3% EO concentration has the potential to induce certain morphological changes in the wings, in contrast to lower concentrations of 1.5%, 0.75%, 0.38%, and 0.19% that were used in previous studies. For the control group, a feeding medium without essential oils was used. Then, twenty larvae per replicate were transferred onto each treatment as well as the control feeding medium. Experimental groups were incubated under standard laboratory conditions, as explained in the previous paragraph of this section. At the end of the fruit flies' life cycle, i.e., after the eclosion of adults (imago), all successfully emerged males and females were collected and preserved in 70% ethanol prior to further analysis.

Wing dissection and slide preparation

Wing dissection from the fruit fly's body was conducted using fine forceps and entomological needles, conducted in small plastic trays filled with distilled water. Detached wings were mounted on microscopic slides using Berlese medium. Details of the process are given in [26]. In each group exposed EOs and in control groups alike, 15 specimens were randomly chosen, and their wings were carefully detached using the method described earlier. Wings were photographed with a Leica Flexicam C3 on a Leica M165 stereomicroscope (Leica Microsystems, Weltzar, Germany) with 40× magnification.

Geometric morphometrics – positioning of landmarks

The assessment of possible variations in *D. melanogaster* wing shape resulting from different EO treatments was based on geometric morphometrics [32]. To depict

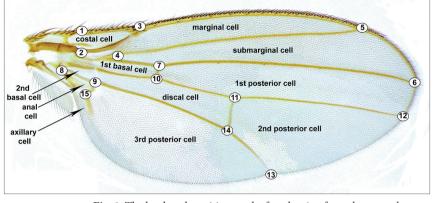


Fig. 1. The landmark position on the female wing from the control group. Fifteen landmarks (LM) depict the shape of the following wing cells: LM 1-3 – costal cell, LM 2-5 – marginal cell, LM 4-6 – submarginal cell, LM 2, 4, 7, 8, and 10 first basal cell, LM 6-7 and LM 10-12 – first posterior cell, LM 8-9 – length of second basal cell, LM 9-11 and LM 14 – discal cell, LM 11-14 – second posterior cell, LM 8-9 and LM 15 – anal cell, LM 9 and LM 13-15 – third posterior cell. Wing cell nomenclature is given according to [35].

the wing shape, 15 landmarks were positioned in the tpsDig2 software [33] following Gidaszewski et al. [34]. All landmarks are placed at the intersection of veins (Fig. 1). Wing cell nomenclature follows Morgan et al. [35]. To eliminate discrepancies in position, rotation, and translation, a generalized Procrustes analysis (GPA) was performed as outlined by Dryden and Mardia [36] and Rohlf and Slice [37]. The shape and size were obtained as Procrustes coordinates and centroid size (CS), respectively. GPA was performed in MorphoJ software [38].

Statistical analysis

The previous experiments, from which the flies used in this study originated, were categorized according to genus affiliation (*Pinus* and *Abies*) and conducted at different times. Differences in the composition of the EOs between these two genera led us to treat them as distinct entities throughout the manuscript and perform separate analyses within the *Pinus* and *Abies* groups. Also, considering that we analyzed EOs from nine species, for practical reasons and readability, we decided to separate the analysis according to genus affiliation. The positioning of males and females in the morphospace suggests that any changes in wing morphology resulting from exposure to EOs were smaller compared to the inherent differences in wing

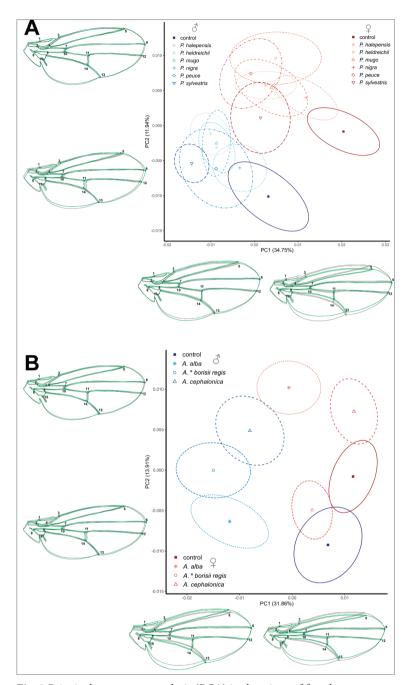


Fig. 2 Principal component analysis (PCA) in the wings of females and males exposed to the essential oils of 6 *Pinus* species (**A**) and 3 *Abies* species (**B**) and their corresponding control groups. The morphospace was constructed using the first two PC axes. EO treatments are shown as ellipses with dashed lines and assigned symbols; controls are in a solid line. Females are represented by shades of red, and males in shades of blue. The ellipses represent a 90% confidence interval for group means. Changes in wing morphology along the PC axes are shown via contour graphs; grey lines show the mean shape of the wing, and green lines describe the changes in the maximum or minimum value for both axes.

shape and size between males and females in *D. melanogaster*. Therefore, separate analyses were conducted within the male and female groups for both *Abies* and *Pinus* species' EOs.

To assess the differences in wing size, analysis of variance (ANOVA) was conducted using the logarithmic value of centroid size (logCS) as the dependent variable. The factors considered were sex, treatment, and their interaction. The specific tests included differences in wing size between sexes (logCS ~ sex), differences in wing size among treatments (logCS ~ treatment), and interaction effects of sex and treatment on wing size (logCS ~ sex * treatment). For analyzing wing shape, multivariate analysis of variance (MANOVA) was applied to Procrustes coordinates, focusing on the following factors: shape differences between sexes (shape ~ sex), shape differences among treatments (shape ~ treatment), interaction effects on shape between sex and treatment (shape ~ sex * treatment). All statistical analyses were performed using the geomorph package version 4.0.5. [39] in R Studio software [40]. Significant differences were considered as P<0.05. Spearman's rank correlation coefficient (Spearman's Rho) was used to analyze correlations between the selected compounds, constituents of the tested essential oils (EOs) (Supplementary Table S1), and the centroid size value (which describes wing size). These analyses were performed separately within Pinus and Abies species, as well as separately for females and males. The correlation analysis was done in software SPSS version 15.0 (SPSS Inc, Chicago, IL, USA). Principal component analysis (PCA) and canonical variate analyses (CVA) were used

for testing wing divergence. Both PCA and CVA were performed in the MorphoJ software [38]. The results of PCA and CVA were extracted and used for generating scatterplot graphs in R Studio software [40] via the ggplot2 software package [41]. Additionally, within the CVA, the Procrustes distances were calculated as a value of deviations in the wing shape of treated flies from the control group.

RESULTS

Principal component analysis (PCA) revealed distinct patterns of wing shape variation between the control and treated groups, as well as between sexes. In both females and males, discrimination between control groups was evident along the first principal component (PC1) axis, whereas differentiation between sexes was observed along the second principal component (PC2) axis (Fig. 2A). The discrepancy between the control group and the specimens exposed to essential oils mainly pertained to the width of the wings. Female controls

exhibited the narrowest wings, whereas wings in the

treated groups and male controls were similar in shape. Notably, the widest wings were found in the male groups exposed to EOs, with the wings of those exposed to the *P. sylvestris* EO being the widest compared to the male control group. Among female groups, the greatest difference was between wings exposed to the EO of *P. peuce* and the control group (Fig. 2A). Small differences along the PC2 axis (PC2 = 11.94%) explained sex-related differences, with females exhibiting slightly wider and shorter wings with shorter first and second posterior cells. In contrast, males showed elongated and narrower wings with longer first and second posterior cells (Fig. 2A).

ANOVA indicated statistically significant differences in wing size (Z = 5.899, P = 0.001) and in the interaction between sex and treatment

(Z = 2.111, P = 0.018). However, differences in wing size between sexes and between treatments alone were not statistically significant. The multivariate analysis of variance (MANOVA) revealed statistically significant differences in wing shape between sexes (Z = 12.940, P = 0.001), among treatments (Z = 6.343, P = 0.001), and in the interaction between sex and treatment (Z = 1.703, P = 0.047).

Due to the statistically significant results of ANOVA and MANOVA, CVA was applied. The sexual dimorphism in wing morphology was sustained even after exposure of *D. melanogaster* larvae to the essential oils (Fig. 3). Notably, all female groups, including the control, clustered in the negative part of the CV1 axis, while all males, along with with their respective control, clustered in the positive part of CV1. Despite

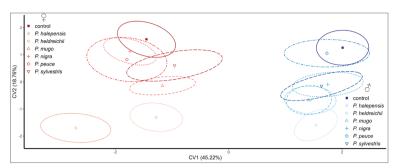


Fig. 3. Comparative analysis of changes in the wings of females and males exposed to the essential oils of six *Pinus* species and their control groups. The morphospace was constructed using the first two CV axes. EO treatments are shown as ellipses with dashed lines and assigned symbols; controls are in a solid line. Females are represented by shades of red and males in shades of blue. The ellipses represent a 90% confidence interval for group means.

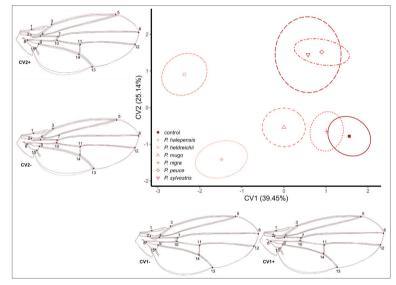


Fig. 4. Variation of females' wings exposed to essential oils of six *Pinus* species and control group in CV1 \times CV2 morphospace. EO treatments are shown as ellipses with dashed lines and assigned symbols; controls are in a solid line. Changes in wing morphology along the CV axes are shown via contour graphs; grey lines show the mean shape of the wing, and red lines describe the changes in the maximum or minimum value for both axes. The ellipses represent a 90% confidence interval for group means.

the influence of the *Pinus* EOs on wing morphology, sexual dimorphism is evident. This fact eliminates any concerns about mixing groups of different sexes, which is essential for the subsequent interpretation of the results when analyzing the wing changes separately in females and males (Figs. 4 and 5.)

The total variability of wing morphology summarizing CV1+CV2 was 64.59%. The most noticeable separation along the CV1 axis is demonstrated

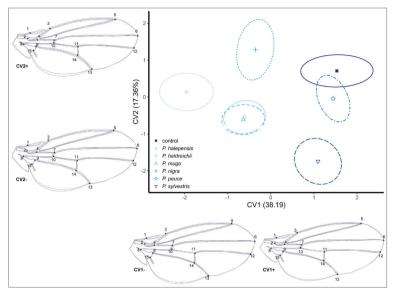


Fig. 5. Variation of male wings exposed to essential oils of six *Pinus* species and control group in $CV1 \times CV2$ morphospace. EO treatments are shown as ellipses with dashed lines and assigned symbols; controls are in a solid line. The changes in wing morphology along the CV axes are presented in contour graphs. Grey lines show the mean shape of the wing, and blue lines describe the changes in the maximum or minimum value for both axes. The ellipses represent a 90% confidence interval for groups' means.

by the P. halepensis and P. heldreichii groups (Fig. 4). CV1 explains 39.45% of the overall variability in wing shape. The control group of females is positioned opposite the P. halepensis and P. heldreichii treatments and is located at the extreme range of the positive part of the CV1 axis. Changes in wing morphology, as observed on the deformation grid, were attributed to alterations in the following wing cells: the third posterior cell, first basal cell, and discal cell. Parallel elongation of the wings results from the shortening of the first and second posterior cells. Along the CV2, which accounts for about 25% of the total wing variability, distinct morphological differences are evident in the treatments involving P. heldreichii, P. peuce and P. sylvestris compared to the control group and the other three treatments: P. halepensis, P. nigra, and P. mugo. Changes in wing shape along the CV2 axis are similar to those observed along the CV1. Wings of specimens exposed to the EOs of P. heldreichii, P. peuce, and P. sylvestris are generally wider, with wider submarginal and second posterior cells (Fig. 4). The P. nigra treatment overlaps significantly with the control group, although the effect of the essential oils of this pine species on wing shape is minimal.

In the male groups (Fig. 5), within the two-dimensional morphospace defined by CV1 and CV2, which together account for 55.55% of the total wing variability, the control group was located in the $CV1(+) \times CV2(+)$ quadrant. The other treatments are scattered throughout the CVA morphospace, indicating that each treatment induces distinct changes in *D. melanogaster* wing shape, except for *P.* mugo and P. heldreichii, which overlapAlong the first canonical axis, which accounts for 38.19% of the total wing morphology variability, the treatments P. peuce and P. sylvestris are positioned in the positive part of CV1, while the other four treatments, P. halepensis, P. heldreichii, P. nigra, and P. mugo, are placed in the opposite part (Fig. 5). In these four

treatments, changes can be observed in the form of an expansion of the entire wing, primarily the third posterior cell. Slight changes were observed, primarily as a mild lengthening of the first basal cell and a subtle shortening of the first posterior cell of the wing. In comparison to the other treatments, the wings of flies subjected to *P. peuce* EOs displayed the smallest changes in morphology, with considerable overlap with the control group. The second canonical axis, contributing only 17.36% to wing differentiation, polarizes the remaining two EO treatments, *P. nigra* in the CV2 (+) direction and *P. sylvestris* in the CV2 (-) direction. Male wings exposed to *P. sylvestris* EOs have wide second discal and third posterior cells (Fig. 5), in contrast to wings treated with *P. nigra* EOs.

Visualization of the results by PCA revealed clear distinctions in wing shape between the groups exposed to Abies EOs and the control groups. In males, the control group was clearly separated from those treated with EOs along the PC1 axis. For females, the control group differed significantly from the group exposed to A. alba EOs (Fig. 2B). These differences were primarily attributed to variations in wing width. Specifically, control males and females had narrower wings, while males exposed to EOs from the three Abies species had wider wings (Fig. 2B). Along the PC2 axis, the male control and A. alba groups were separated from the A. cephalonica group. In females, exposure to A. \times borisii-regis EO resulted in narrower wings with slightly longer first and second posterior cells compared to the effects observed with A. alba (Fig. 2B).

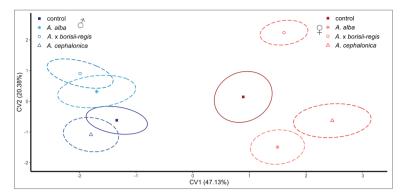


Fig. 6. Comparative analysis of changes in the wings of females and males exposed to the essential oils of three *Abies* species and their control groups. The morphospace was constructed using the first two CV axes. EO treatments are shown as ellipses with dashed lines and assigned symbols; controls are in a solid line. Females are represented by shades of red, and males in shades of blue. The ellipses represent a 90% confidence interval for group means.

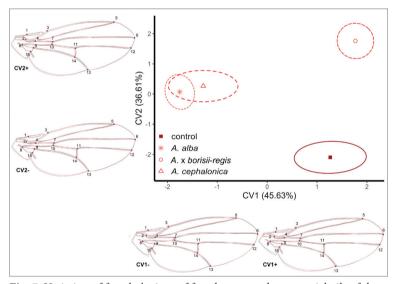


Fig. 7. Variation of females' wings of females exposed to essential oils of three *Abies* species and control group in CV1 \times CV2 morphospace. EO treatments are shown as ellipses with dashed lines and assigned symbols; controls are in a solid line. The changes in wing morphology along the CV axes are presented in contour graphs. Grey lines show the mean shape of the wing, and red lines describe the changes in the maximum or minimum value for both axes. The ellipses represent a 90% confidence interval for group means.

P=0.001), and in the interaction between sex and treatment (Z=1.713, P=0.039).

Similar to the previous analysis of wing differentiation in *Pinus* treatments, Fig. 6 confirms the presence of sexual dimorphism in the *Abies* EO treatments as well. Along the CV1 axis, all female groups, including the control group, are positioned positively, whereas all male groups, including their respective controls, are positioned negatively. The analyses were conducted separately for male and female groups.

In female Abies specimens, both CV axes are crucial for distinguishing individuals based on wing shape and size. CV1, explaining 45.63% of the total variability, separates the control and A. × borisii-regis treatment from the A. alba and A. cephalonica treatments (Fig. 7). Morphological differences in wings between the A. alba and A. cephalonica treatments are minimal. Conversely, the control group is visibly segregated from the A. × borisii-regis treatment along the CV2 axis (describes 36.61% of the total wing variability). Changes in wing shape are reflected in the width and length of the first basal, discal, first, second, and third posterior cells. Wings of specimens exposed to EOs of A. alba and A. cephalonica have a wider third posterior cell, vaguely elongated first basal and discal cells, and marginally shorter first and second posterior cells (Fig. 7). EO of A. × borisii-regis affected the enlargement of the submarginal and second posterior cells, as well as the elongation of the second posterior cell and the reduction of

Results of ANOVA indicated statistically significant differences in wing size (Z=5.3116, P=0.001) and the interaction between wing size and treatment (Z=3.911, P=0.001). However, no significant differences were observed in wing size between the sexes or in the interaction between sex and treatment. MANOVA showed significant differences in wing shape between sexes (Z=8.170, P=0.001) across treatments (Z=3.743,

discal cell length.

In males, there is significant discrimination along CV1 (46.58%) between the control group and those groups exposed to one of the three essential oils, as depicted in Fig. 8. The most distinctive wings, when compared to the wings from the control group, were those exposed to the *A. cephalonica and A.* × *borisiiregis* EOs. The most pronounced differences in wing

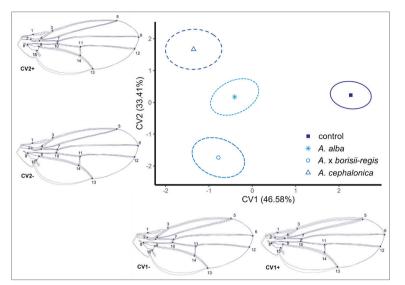


Fig. 8. Variation of males' wings exposed to essential oils of three *Abies* species and control group in $CV1 \times CV2$ morphospace. EO treatments are shown as ellipses with dashed lines and assigned symbols; controls are in a solid line. The changes in wing morphology along the CV axes are presented in contour graphs. Grey lines show the mean shape of the wing, blue lines describe the changes of the maximum or minimum value for both axes. The ellipses represent a 90% confidence interval for group means.

morphology of A. cephalonica are characterized by a general broadening of the wings along the longitudinal axis. These wings have broader discal, second and third posterior, and submarginal cells, along with slightly elongated first basal cells (Fig. 8). On the second canonical axis, which accounts for 33.41% of the total variability, two groups can be discerned. The wings of A. cephalonica are positioned in the positive part, whereas the wings exposed to A. × *borisii-regis* EO are grouped in the negative part of CV2. The main disparities between these two groups are seen in the width of the wing as well as in the width of the discal, marginal, second, and third posterior cells. The wings treated with A. × borisii-regis EO are wider, with all the afore-mentioned cells being wider when compared to those treated with A. cephalonica EO (Fig. 8). The wings from the A. alba treatment are situated between the wings of the A. \times borisii-regis and A. cephalonica treatments.

We employed Procrustes distances as a numerical measure that indirectly indicates changes in wing shapes. This value quantifies the deviations in the wing shapes of treated fruit flies compared to the control (untreated flies). Procrustes distances were separately calculated for females and males, as well as individually within the *Pinus* and *Abies* groups, all presented in Table 1. Procrustes distances were significant in all tested EOs except the EO of *P. nigra* which showed no significant influence on changes in wing shape in both genders, while the *P. peuce* EO did not exhibit significant influence in males. Additionally, in the *Abies* group among females, only the essential oil from *A. cephalonica* did not show a significant influence on wing shape.

Spearman's correlation analysis (Table 2) revealed that wing size is significantly negatively correlated with the concentrations of α -humulene, (*E*)-caryophyllene, and thunbergol in both female and male individu-

als treated with *Pinus* EOs. For α -pinene, a negative correlation with wing size is only observed in males. In individuals treated with *Abies* EOs, wing size was significantly negatively correlated with the concentration of bornyl acetate and germacrene D in both females and males. Additionally, significant negative correlations were found between wing size and the concentration of α -humulene and (*E*)-caryophyllene only in females and limonene+ β -phellandrene and camphene only in males. However, considering that all obtained Rho values < 0.5 (Table 2), these significant correlations can be classified as weak.

DISCUSSION

Plant-derived essential oils have been acknowledged as botanical pesticides [42], showing significant potential for integration into organic crop production as alternatives to synthetic chemical pesticides, which may pose risks to both humans and the environment [43]. They have demonstrated insecticidal and repellent activities against mosquitoes [27,44] and insecticidal and deterrent effects against *D. suzukii*, which is an important invasive fruit pest [23,24]. In pursuit of this, frequently examined components found in EOs include thymol, thujone, carvacrol, and limonene, and there are even commercial products based on EOs [42].

At a concentration of 3%, *P. sylvestris* and *P. peuce* EOs induced significantly higher larvicidal activity and delayed developmental time in *D. melanogaster* compared to *P. nigra* and *P. mugo* EOs, respectively [4]. Additionally, the EO of *P. heldreichii* also exhibited

		Pinus group							Abies group		
Treatment		P. halepensis	P. heldreichii	P. mugo	P. nigra	P. peuce	P. sylvestris	A. alba	A. borisii × regis	A. cephalonica	
	females	0.0212***	0.0206**	0.0182**	0.0119	0.0238***	0.0207**	0.0189***	0.0139*	0.0114	
sex	males	0.019***	0.0164**	0.0169**	0.0126	0.0147	0.021**	0.0204**	0.0258***	0.0214***	

Table 1. Deviations of the wing shape of the treatment from the control (Procrustes distances) separately presented for females and males

*P<0.05; **P<0.01; ***P<0.001

Table 2. Correlation of the components from three *Abies* and six *Pinus* EOs with centroid size (describing wings size) separately for females and males

Compounds from the	Pinu	s EOs	Abies EOs		
tested EO	male	female	male	female	
α-pinene	-0.204*	-0.122	-0.073	0.133	
β-pinene	-0.145	-0.075	-0.073	0.133	
α-humulene	-0.317**	-0.410**	-0.240	-0.389**	
limonene+β-phellandrene	0.065	0.093	-0.334*	-0.101	
camphene	-0.103	-0.014	-0.334*	-0.101	
bornyl acetate	-0.122	-0.037	-0.437**	-0.362**	
germacrene D	-0.125	-0.120	-0.272*	-0.452**	
β -caryophyllene	-0.250*	-0.336**	-0.240	-0.389**	
thunbergol	-0.221*	-0.270*	n/a	n/a	
δ-3-Carene	-0.094	-0.101	-0.199	-0.166	

* P<0.05; ** P<0.01; "n/a" – not applicable due to lack of the compound within the tested EO within genera

larvicidal effect at 3% against *D. melanogaster* [5]. However, changes in wing morphology do not exactly align with the reported larvicidal activity trend. For instance, in the female group, the impact on wing morphology was induced in the following order by EOs: P. halepensis, P. heldreichii, P. peuce, P. sylvestris, P. mugo, and P. nigra, from strongest to weakest, respectively. Interestingly, the EOs of P. heldreichii, P. peuce, and P. sylvestris had qualitatively different effects on wing morphology compared to treatments with P. halepensis, P. nigra, and P. mugo. These differences might be related to the previously reported larvicidal activity of the mentioned EOs against D. melanogaster [4,5]. Apart from *P. halepensis*, α -pinene was one of the first two major compounds in all the studied Pinus EOs, and its content was positively correlated to the inhibition of developmental time and larvicidal activity [4,5]. In addition, α -pinene is a well-known compound with bioactivity against insect pests, including fumigant toxicity against Sitophilus oryzae [45], Plutella xylostella, and Megoura japonica [46], while also affecting the reproduction of the green peach aphid, Myzus persicae [47]. However, a high relative percentage content of pinenes in Pinus EOs did not correspond to the same intensity of changes in wing morphology changes. Notably, the EOs of P. mugo and P. nigra previously showed no toxicity or very weak toxicity against D. melanogaster, respectively [4]. Despite a high relative percentage content of pinenes, P. nigra EO had an insignificant impact on wing shape in both males and females, unlike the other tested Pinus EOs. The effect on female and male flies treated with P. mugo EO resulted in significant differences in wing shape (Table 1). Although δ -3carene, which was present in a higher percentage in P. mugo, has been reported to have synergistic activity with selected insecticides [48], it evidently may contributed to this observation. In contrast to females, P. peuce EO had an insignificant effect on changes in wing shape in males, while P. sylvestris, like its impact on females, had a moderate but sig-

nificant effect on changes of wing shape. As in females, P. sylvestris EO had a qualitatively different impact on wing morphology in males compared to P. halepensis, P. heldreichii, P. nigra, and P. mugo. This observation may be attributed to the different composition of P. sylvestris EO compared to other Pinus EOs. In addition, a significant negative correlation between wing size and α -pinene was found in the groups treated with Pinus EOs. Due to its activity against insect pests and vectors, the EO from P. sylvestris is already a candidate for potential use as an insecticide [49]. In both males and females, P. heldreichii EO exhibited a strong effect on changes in wing morphology, especially in shape. *Pinus heldreichii* EO apart from a high α -pinene content, contained higher contents of germacrene D and limonene + β -Phellandrene than the EOs of *P. peuce*, *P.* sylvestris, and P. mugo. These components might affect only shape because they are not significantly correlated with wing size. A recent study has shown the toxic effects of germacrene D on third-instar larvae of insects [50]. Similarly, limonene has induced morphological and physiological deformities in D. suzukii [51], which aligns with the stronger impact of P. heldreichii EO

on wing shape in this study. The P. halepensis EO had the strongest effect on wing morphology, especially shape, in both sexes, despite its very low content of pinenes and the absence of larvicidal activity in D. melanogaster [5]. P. halepensis EO was distinct in its high relative percentage content of thunbergol and β -caryophyllene, unlike the other investigated pine essential oils where a-pinene was consistently one of the two major compounds. While there are no specific toxicological studies on thunbergol for direct comparisons, β -caryophyllene showed potential as an anti-insecticidal agent, impacting the reproduction of *M. persicae* [47] and demonstrating fumigant toxicity against Sitophilus oryzae [45]. Furthermore, β -caryophyllene derived from *Cephalotaxus sinensis* showed the highest contact toxicity against M. japonica and P. xylostella [46]. This suggests that pinenes, known for their larvicidal and toxic effects on pest insects, may not be the primary drivers of wing morphology changes in D. melanogaster. Hence, EOs with a high relative percentage content of other components, such as β -caryophyllene and thunbergol or limonene combined with β -phellandrene and germacrene D, could play a more significant role. Consistent with this conclusion, a statistically significant negative correlation between wing size and the concentration of β -caryophyllene and thunbergol was found in both female and male groups. A noteworthy finding is that the variation of minor component, particularly α -humulene, was also found to be significantly negatively correlated with wing size in both females and males treated with Pinus EOs.

The literature data regarding the effects of Abies EOs on insects is limited [6,48]. Recent studies revealed that Abies balsamea EO exhibited potential as a potent synergist for select insecticides, suggesting its use as an additive for insecticidal formulations [48]. Significant differences in wing morphology were observed in both female and male D. melanogaster flies treated with Abies EOs. Male flies exhibited more pronounced and significant changes in wing shape changes when treated with A. cephalonica and A. × borisii-regis EOs. Abies alba had a milder effect on wing morphology but still showed a significantly difference in wing shape compared to the control. This trend aligns with the toxicity of Abies EOs in D. melanogaster at a 3% concentration, where A. × borisii-regis and A. cephalonica exhibited higher activity than A. alba EO [6]. The major volatiles in all three EOs were β -pinene and α -pinene, with higher levels in A. cephalonica than in the other two species, A. alba and A. \times borisii-regis. These components are known for their detrimental effect on insects, which might influence the wing morphology in D. melanogaster. For instance, β -pinene has repellent activity and affects neurotransmission-related genes in the red flour beetle Tribolium castaneum [52]. EO isolated from *Haplophyllum dauricum*, rich in α -pinene and β -pinene, displayed contact and fumigant toxicity, as well as repellent activity against two pest insects [53]. In addition, α -pinene exhibited an array of toxic activities against pest insects [45-47]. Abies × borisii*regis* was the richest in β -caryophyllene compared to the other two *Abies* species. The bioactivity of β -caryophyllene in insects was discussed in the preceding paragraph. Nevertheless, it is noteworthy that A. cephalonica and A. × borisii-regis EOs induced distinct qualitative changes in male wing shape, which could be attributed to significant differences in the relative percentage content of α -pinene and β -pinene. In addition, values of limonene + β -Phellandrene, bornyl acetate and germacrene D are significantly negatively correlated with wing size in males.

In the female group, A. alba EO induced the greatest changes in wing shape, followed by A. × borisii-regis, while A. cephalonica induced minor differences in wing shape compared to the control and the other two species. Camphene and limonene + β -phellandrene were the third and fourth most abundant components in all three Abies EOs, with a higher percentage in A. alba. As previously mentioned, limonene is considered a promising bioinsecticide that causes morphological and physiological deformities in the well-known pest insect D. suzukii larvae [51], which is concurrent with the results of this study. In addition, (R)-(+)-limonene acts as a repellent against D. melanogaster and D. suzukii [54], while (-)-limonene exhibits strong contact and/or fumigant toxicities against S. oryzae and T. castaneum [55]. Additionally, A. alba essential oil exhibited the highest content of bornyl acetate and camphene compared to the other two Abies species, indicating that these components may play a significant role in inducing changes in wing morphology, either in conjunction with or independently from pinenes. Furthermore, a significant negative correlation was found between wing size and these two compounds within Abies essential oils. Camphene and bornyl acetate have demonstrated fumigant and contact toxicity against storage pest insects

[56]. Camphene has also shown insecticidal activity [57], including fumigant and contact toxicity against pests like the palm weevil Rhynchophorus ferrugineus [58], as well as larvicidal and insecticidal activities. [59]. In females, the induced wing morphological changes were small between A. alba and A. cephalonica but qualitatively different in in terms of width and length compared to $A. \times borisii$ -regis. This could be due to the significant difference in relative percentage content of the major constituents pinenes, limonene + β -phellandrene, and bornyl acetate. The concentrations of β -caryophyllene, α -humulene, and germacrene D within *Abies* EOs are significantly negatively correlated with wing size. There are limited studies on α -humulene insecticidal activity [57] and none for its influence on wing morphology in insects. The observed differences in the response to the same treatment between males and females seen in this study have also been recorded in previous studies [26], and are likely due to physiological differences between the two sexes influencing their responses to the same treatment. Likewise, in treatments involving Pinus EOs, components such as limonene + β -phellandrene, camphene, bornyl acetate, and β -caryophyllene may play an important role in inducing wing morphology changes, either in conjunction with or separately from pinenes. Additionally, it is possible that the combined action of minor components, such as α -humulene and germacrene D, with other components within the Abies EOs contributes to the observed changes in D. melanogaster wing morphology.

CONCLUSIONS

The geometric morphometrics method demonstrates its potential for screening minute morphological changes in *D. melanogaster* wings induced by EO treatments. These changes can be correlated with EO treatment toxicity and the specific chemical composition of each tested EO. At the applied concentration of 3%, the *Abies* and *Pinus* EOs did not cause changes in wing morphology that would impair the wing function of flies. However, almost all recorded changes were significant compared to the control group, indicating a potential to cause morphological changes in wing structure. In both male and female flies, it was observed that EOs derived from *P. halepensis* showed the most pronounced influence on wing morphology, while *P. nigra* EO had the lowest impact, particularly

insignificant for wing shape. EOs from P. heldreichii and P. sylvestris exhibited moderate influence, followed by a moderate to lower impact by the P. mugo EO. In males, P. peuce EOs had none to lower impact and vice versa in females. The A. × borisii-regis EO exhibited the strongest influence in the male group, A. alba EO in the female group, while the A. cephalonica EO exhibited moderate but significant impact on wing shape in males but not in females. Pinenes may not be the primary and necessary factors driving changes in wing morphology in D. melanogaster. Other components, such as β -caryophyllene, thunbergol, limonene + β -phellandrene, camphene, bornyl acetate, or minor ones such as α -humulene and germacrene D, could play a more significant role in inducing changes in D. melanogaster wing morphology. Considering that the dietary treatments caused pronounced morphological changes in D. melanogaster wings, it could be assumed that EOs from the six Pinus and three Abies species might also have the potential to cause changes in other body parts of the insects. We believe that this study lays the foundation for further investigations into the potential of selected Abies and Pinus EOs as biopesticides for insect pest control, with a focus on their capacity to disrupt certain activities that depend on the morphology of the wings, potentially incapacitating or significantly reducing pest insect activities and behaviors. Naturally, these suggestions should be verified in targeted toxicological investigations against pest insects.

Funding: This work was funded by the Ministry of Science, Technological Development and Innovations of the Republic of Serbia [Grant Nos.: 451-03-65/2024-03/200124 and 451-03-66/2024-03/200124].

Author contributions: Conceptualization, VJC; methodology, VJC, ML, VŽ; validation, VJC, M.L., VŽ; formal analysis ML and MSP; investigation, VJC, ML, VŽ; resources, VJC, ZSM, BZ, SJ, GS, VŽ; data curation, ML; writing – original draft preparation, VJC, ML, VŽ; writing – review and editing, VJC, ML, ZSM, BZ, MSP, SJ, GS, VŽ; visualization, ML, VŽ; supervision, VŽ; project administration, VŽ; funding acquisition, VŽ. All authors have read and agreed to the published version of the manuscript.

Conflict of interest disclosure: None of the authors have a conflict of interest to disclose.

Data availability: Data underlying the reported findings have been provided as a raw dataset available here: https://www.serbiosoc.org. rs/NewUploads/Uploads/Cvetkovic%20et%20al_9901_Raw%20 Dataset.docx

REFERENCES

- WFO: World Flora Online [Internet]. c2023 [cited 2023 Jan 9]. Available from: http://www.worldfloraonline.org.
- Menković N, Šavikin K, Tasić S, Zdunić G, Stešević D, Milosavljević S, Vincek D. Ethnobotanical study on traditional uses of wild medicinal plants in Prokletije Mountains (Montenegro). J Ethnopharmacol. 2011;133:97-107. https://doi.org/10.1016/j.jep.2010.09.008
- Yang S-A, Jeon S-K, Lee E-J, Im N-K, Jhee K-H, Lee S-P, Lee I-S. Radical scavenging activity of the essential oil of Silver fir (*Abies alba*). J Clin Biochem Nutr. 2009;44(3):253-259. https://doi.org/10.3164/jcbn.08-240
- Mitić ZS, Jovanović B, Jovanović SČ, Mihajilov-Krstev T, Stojanović-Radić ZZ, Cvetković VJ, Mitrović TL, Marin PD, Zlatković BK, Stojanović GS. Comparative study of the essential oils of four *Pinus* species: Chemical composition, antimicrobial and insect larvicidal activity. Ind Crops Prod. 2018;111:55-62. https://doi.org/10.1016/j.indcrop.2017.10.004
- Mitić ZS, Jovanović B, Jovanović SČ, Stojanović-Radić ZZ, Mihajilov-Krstev T, Jovanović NM, Nikolić BM, Marin PD, Zlatković BK, Stojanović GS. Essential oils of *Pinus halepensis* and *P. heldreichii*: Chemical composition, antimicrobial and insect larvicidal activity. Ind Crops Prod. 2019;140:111702. https://doi.org/10.1016/j.indcrop.2019.111702
- Mitić ZS, Stojanović-Radić ZZ, Jovanović SČ, Cvetković VJ, Nikolić JS, Ickovski JD, Mitrović TL, Nikolić BM, Zlatković BK, Stojanović GS. Essential oils of three Balkan Abies species: Chemical profiles, antimicrobial activity and toxicity toward Artemia salina and Drosophila melanogaster. Chem Biodivers. 2022;19(6):e202200235. https://doi.org/10.1002/cbdv.202200235
 - Lahlou M. Composition and mollussiside
- Lahlou M. Composition and molluscicidal properties of essential oils of five Moroccan Pinaceae. Pharm Biol. 2003; 41:207-210. https://doi.org/10.1076/phbi.41.3.207.15097
- Koutsaviti K, Giatropoulos A, Pitarokili D, Papachristos D, Michaelakis A, Tzakou O. Greek Pinus essential oils: larvicidal activity and repellency against *Aedes albopictus* (Diptera: Culicidae). Parasitol Res. 2015;114:583-592. https://doi.org/10.1007/s00436-014-4220-2
- Govindarajan M, Rajeswary M, Benelli G. Chemical composition, toxicity and non-target effects of *Pinus kesiya* essential oil: An eco-friendly and novel larvicide against malaria, dengue and lymphatic filariasis mosquito vectors. Ecotoxicol Environ Saf. 2016;129:85-90.

https://doi.org/10.1016/j.ecoenv.2016.03.007

 Wajs-Bonikowska A, Sienkiewicz M, Stobiecka A, Maciąg A, Szoka Ł, Karna E. Chemical composition and biological activity of *Abies alba* and *A. koreana* seed and cone essential oils and characterization of their seed hydrolates. Chem Biodivers. 2015;12(3):407-18.

https://doi.org/10.1002/cbdv.201400167

 Xie Q, Liu Z, Li Z. Chemical composition and antioxidant activity of essential oil of six *Pinus* taxa native to China. Molecules. 2015;20(5):9380-92. https://doi.org/10.3390/molecules20059380

https://doi.org/10.3390/molecules20059380

12. Tognolini M, Barocelli E, Ballabeni V, Bruni R, Bianchi A, Chiavarini M, Impicciatore M. Comparative screening of plant essential oils: phenylpropanoid moiety as basic core for antiplatelet activity. Life Sci J. 2006;78(13):1419-32. https://doi.org/10.1016/j.lfs.2005.07.020

- Hoai NT, Duc HV, Thao DT, Orav A, Raal A. Selectivity of *Pinus sylvestris* extract and essential oil to estrogen-insensitive breast cancer cells. Pharmacogn Mag. 2015;11(44s1):S290-5. https://doi.org/10.4103/0973-1296.166052
- 14. Roberts DB. *Drosophila melanogaster*: the model organism. Entomologia experimentalis et applicata. 2006;121(2):93-103. https://doi.org/10.1111/j.1570-8703.2006.00474.x
- Moraes KC, Montagne J. Drosophila melanogaster: A powerful tiny animal model for the study of metabolic hepatic diseases. Front Physiol. 2021;12:728407. https://doi.org/10.3389/fphys.2021.728407
- Žabar A, Cvetković V, Rajković J, Jović J, Vasiljević P, Mitrović T. Larvicidal activity and *in vitro* effects of green tea (*Camellia sinensis* L.) water infusion. Biol Nyssana. 2013;4:75-9.
- Franzios G, Mirotsou M, Hatziapostolou E, Kral J, Scouras ZG, Mavragani-Tsipidou P. Insecticidal and genotoxic activities of mint essential oils. J Agric Food Chem. 1997;45(7):2690-4. https://doi.org/10.1021/jf960685f
- Mihajilov-Krstev T, Jovanović B, Jović J, Ilić B, Miladinović D, Matejić J,Rajković J, Đorđević Lj, Cvetković V, Zlatković B. Antimicrobial, antioxidative, and insect repellent effects of *Artemisia absinthium* essential oil. Planta Med. 2014;80(18):1698-705.

https://doi.org/10.1055/s-0034-1383182

- Mihajilov-Krstev T, Jovanović B, Zlatković B, Matejić J, Vitorović J, Cvetković V, Ilić B, Đorđević L, Joković N, Miladinović D, Jakšić T. Phytochemistry, toxicology and therapeutic value of *Petasites hybridus* subsp. *ochroleucus* (common Butterbur) from the Balkans. Plants. 2020;9(6):700. https://doi.org/10.3390/plants9060700
- Vitorović J, Joković N, Radulović N, Mihajilov-Krstev T, Cvetković VJ, Jovanović N, Mitrović T, Aleksić A, Stanković N, Bernstein N. Antioxidant activity of hemp (*Cannabis* sativa L.) seed oil in *Drosophila melanogaster* larvae under non-stress and H₂O₂-induced oxidative stress conditions. Antioxidants. 2021;10(6):830. https://doi.org/10.3390/antiox10060830

 Bongiorni S, Arisi I, Ceccantoni B, Rossi C, Cresta C, Castellani S, Forgione I, Rinalducci S, Muleo R, Prantera G. Apple polyphenol diet extends lifespan, slows down mitotic

- Apple polyphenol diet extends lifespan, slows down mitotic rate and reduces morphometric parameters in *Drosophila melanogaster*: A comparison between three different apple cultivars. Antioxidants. 2022;11(11):2086. https://doi.org/10.3390/antiox11112086
- Eben A, Sporer F, Vogt H, Wetterauer P, Wink M. Search for alternative control strategies of *Drosophila suzukii* (Diptera: Drosophilidae): Laboratory assays using volatile natural plant compounds. Insects. 2020;11(11):811. https://doi.org/10.3390/insects11110811
- Erland LA, Rheault MR, Mahmoud SS. Insecticidal and oviposition deterrent effects of essential oils and their constituents against the invasive pest *Drosophila suzukii* (Matsumura) (Diptera: Drosophilidae). Crop Prot. 2015;78:20-26. https://doi.org/10.1016/j.cropro.2015.08.013
- 24. Gullickson M, Flavin Hodge C, Hegeman A, Rogers M. Deterrent effects of essential oils on spotted-wing *Drosophila*

(*Drosophila suzukii*): Implications for organic management in berry crops. Insects. 2020;11(8):536.

https://doi.org/10.3390/insects11080536

- Mitić ZS, Stojanović-Radić Z, Cvetković VJ, Jovanović SČ, Dimitrijević M, Ickovski JD, Jovanović N, Mihajilov-Krstev T, Stojanović GS. Pseudotsuga menziesii (Pinaceae): Volatile profiles, antimicrobial activity and toxicological evaluation of its essential oil. Chem Biodivers. 2021;18(9):e2100424. https://doi.org/10.1002/cbdv.202100424
- Cvetković VJ, Jovanović B, Lazarević M, Jovanović N, Savić-Zdravković D, Mitrović T, Žikić V. Changes in the wing shape and size in *Drosophila melanogaster* treated with food grade titanium dioxide nanoparticles (E171) a multigenerational study. Chemosphere. 2020;261:127787. https://doi.org/10.1016/j.chemosphere.2020.127787
- Silva PB, Santos RB, da Cruz RC, da Silva DC, da Silva PS. Effect of *Croton tetradenius* essential oil on larval viability, pupal viability, and wing geometric morphometrics of *Aedes aegypti*. Biocatal Agric Biotechnol. 2023;51:102743. https://doi.org/10.1016/j.bcab.2023.102743
- Lazarević M, Kavallieratos NG, Nika EP, Boukouvala MC, Skourti A, Žikić V, Papanikolaou NE. Does the exposure of parental female adults of the invasive *Trogoderma granarium* Everts to pirimiphos-methyl on concrete affect the morphology of their adult progeny? A geometric morphometric approach. Environ Sci Pollut Res. 2019;26:35061-70. https://doi.org/10.1007/s11356-019-06120-y
- Boukouvala MC, Kavallieratos NG, Žikić V, Štanković SS, Ilić Milošević M, Skourti A, Lazarević M. Sub-lethal effects of pirimiphos-methyl are expressed to different levels in wings of three stored-product coleopterans: A geometric morphometrics investigation. Insects. 2023;14(5):430. https://doi.org/10.3390/insects14050430
- Lorenz C, Almeida F, Almeida-Lopes F, Louise C, Pereira SN, Petersen V, Vidal PO, Virginio F, Suesdek L. Geometric morphometrics in mosquitoes: what has been measured? Infect Genet Evol. 2017;54:205-15. https://doi.org/10.1016/j.meegid.2017.06.029
- Mitić ZS, Jovanović SČ, Zlatković BK, Nikolić BM, Stojanović GS, Marin PD. Needle terpenes as chemotaxonomic markers in *Pinus*: subsections *Pinus* and *Pinaster*. Chem Biodivers. 2017;14(5):e1600453. https://doi.org/10.1002/cbdv.201600453
- 32. Zelditch Ml, Swiderski DL, Sheets HH. Geometric morphometrics for biologists: a primer. 2nd ed. Elsevier Academic Press; 2012.
- Rohlf, F. J. tpsDig, Digitize landmarks and outlines [Software]. Stony Brook, NY: Department of Ecology and Evolution, State University of New York. 2021; [cited 2023 Aug 28]. Available from: https://www.sbmorphometrics.org/soft-dataacq.html.
- Gidaszewski NA, Baylac M, Klingenberg CP. Evolution of sexual dimorphism of wing shape in the *Drosophila melanogaster* subgroup. BMC Evol Biol. 2009;9:110. https://doi.org/10.1186/1471-2148-9-110
- Morgan TH, Bridges CB, Sturtevant AH. Contributions to the Genetics of *Drosophila Melanogaster*.. Washington: Carnegie Institution of Washington; 1919. 388 p. (No. 278)
- Dryden IL, Mardia KV. Statistical shape analysis: Wiley series in probability and statistics. Chichester: John Wiley & Sons; 1998.

- Rohlf FJ, Slice D. Extensions of the Procrustes method for the optimal superimposition of landmarks. Syst Biol.1990;39(1):40-59. https://doi.org/10.2307/2992207
- Klingenberg CP. MorphoJ: an integrated software package for geometric morphometrics. Mol Ecol Resour. 2011;11(2):353-7. https://doi.org/10.1111/j.1755-0998.2010.02924.x
- Adams D, Collyer M, Kaliontzopoulou A, Baken E. Geomorph: software for geometric morphometric analyses [Software]. University of New England. 2023; [cited 2023 Aug 28]. Available from: https://cran.r-project.org/package=geomorph.
- 40. Rstudio Team. Rstudio: Integrated development for R [Software]. PBC, Boston, MA. 2020; [cited 2023 Aug 28]. Available from: http://www.rstudio.com/.
- Wickham H. Ggplot2: Elegant graphics for data analysis. 2nd ed. Cham, Switzerland: Springer International Publishing; 2016. 260 p.
- 42. Campos EV, Proença PL, Oliveira JL, Bakshi M, Abhilash PC, Fraceto LF. Use of botanical insecticides for sustainable agriculture: Future perspectives. Ecol Indic. 2019;105:483-95. https://doi.org/10.1016/j.ecolind.2018.04.038
- 43. Rattan RS. Mechanism of action of insecticidal secondary metabolites of plant origin. Crop Prot. 2010;29(9):913-20. https://doi.org/10.1016/j.cropro.2010.05.008
- de Souza MA, Da Silva L, Macêdo MJF, Lacerda-Neto LJ, dos Santos MAC, Coutinho HDM, Cunha FAB. Adulticide and repellent activity of essential oils against *Aedes aegypti* (Diptera: Culicidae) - a review. South Afr J Bot. 2019;124:160-5. https://doi.org/10.1016/j.sajb.2019.05.007
- Chaubey MK. Fumigant toxicity of essential oils and pure compounds against *Sitophilus oryzae* L. (Coleoptera: Curculionidae). Biol Agric Hortic. 2012;28:111-9. https://doi.org/10.1080/01448765.2012.681352
- Ma S, Jia R, Guo M, Qin K, Zhang L. Insecticidal activity of essential oil from *Cephalotaxus sinensis* and its main components against various agricultural pests. Ind Crops Prod. 2020;150:112403.

https://doi.org/10.1016/j.indcrop.2020.112403

- Chohan TA, Chohan TA, Mumtaz MZ, Alam MW, Naseer I, Riaz A, Naseem T, Iftikhar A, Najaf Ali DE, Hassan M, Ali HM. Insecticidal potential of α-Pinene and β-Caryophyllene against *Myzus persicae* and their impacts on gene expression. Phyton. 2023;92(7):1943-54. https://doi.org/10.32604/phyton.2023.026945
- Norris EJ, Bloomquist JR. Fir (*Abies balsamea*) (Pinales: Pinaceae) needle essential oil enhances the knockdown activity of select insecticides. J Med Entomol. 2023;60(6):1350-6.
- https://doi.org/10.1093/jme/tjad101
 49. Pavela R, Maggi F, Mazzara E, Torresi J, Cianfaglione K, Benelli G, Canale A. Prolonged sublethal effects of essential oils from non-wood parts of nine conifers on key insect pests and vectors. Ind Crops Prod. 2021;168:113590.
 https://doi.org/10.1016/j.indcrop.2021.113590
- Al-Ghanim KA, Krishnappa K, Pandiyan J, Nicoletti M, Gurunathan B, Govindarajan M. Insecticidal potential of matricaria chamomilla's essential oil and its components (E)-β-Farnesene, germacrene D, and α-Bisabolol oxide A against agricultural pests, malaria, and zika virus vectors. Agriculture. 2023;13(4):779. https://doi.org/10.3390/agriculture13040779

279

- de Souza MT, de Souza MT, Bernardi D, de Melo DJ, Zarbin PHG, Zawadneak MAC. Insecticidal and oviposition deterrent effects of essential oils of *Baccharis* spp. and histological assessment against *Drosophila suzukii* (Diptera: Drosophilidae). Sci Rep. 2021;11:3944. https://doi.org/10.1038/s41598-021-83557-7
- Pajaro-Castro N, Caballero-Gallardo K, Olivero-Verbel J. Neurotoxic effects of linalool and β-pinene on Tribolium castaneum Herbst. Molecules. 2017;22(12):2052. https://doi.org/10.3390/molecules22122052
- 53. Cao JQ, Pang X, Guo SS, Wang Y, Geng ZF, Sang YL, Guo PJ, Du SS. Pinene-rich essential oils from Haplophyllum dauricum (L.) G. Don display anti-insect activity on two stored-product insects. Int Biodeter Biodegr. 2019;140:1-8. https://doi.org/10.1016/j.ibiod.2019.03.007
- 54. Wang Q, Xu P, Sanchez S, Duran P, Andreazza F, Isaacs R, Dong K. Behavioral and physiological responses of *Drosophila melanogaster* and *D. suzukii* to volatiles from plant essential oils. Pest Manag Sci. 2021;77(8):3698-705. https://doi.org/10.1002/ps.6282
- 55. Abdelgaleil SAM, Mohamed MIE, Badawy MEI, El-arami SAA. Fumigant and contact toxicities of monoterpenes to *Sitophilus oryzae* (L.) and *Tribolium castaneum* (Herbst) and their inhibitory effects on acetylcholinesterase activity. J Chem Ecol. 2009;35:518-25. https://doi.org/10.1007/s10886-009-9635-3

SUPPLEMENTARY MATERIAL

- 56. Feng Y-X, Wang Y, Chen Z-Y, Guo S-S, You C-X, Du S-S. Efficacy of bornyl acetate and camphene from *Valeriana* officinalis essential oil against two storage insects. Environ Sci Pollut Res. 2019;26:16157-65. https://doi.org/10.1007/s11356-019-05035-y
- 57. Benelli G, Govindarajan M, Rajeswary M, Vaseeharan B, Alyahya SA, Alharbi NS, Kadaikunnan S, Khaled JM, Maggi F. Insecticidal activity of camphene, zerumbone and α-Humulene from *Cheilocostus speciosus* rhizome essential oil against the old-world bollworm, *Helicoverpa armigera*. Ecotoxicol. Environ Saf. 2018;148:781-6.

https://doi.org/10.1016/j.ecoenv.2017.11.044

 Sharaby A, El-Dosary M. Possibility using camphene as biorational insecticide against the red palm weevil *Rhyn-chophorus ferrugineus* (Coleoptera: Curculionedae). Int J Sci Res. 2016;5:222-5.

https://doi.org/10.21275/ART2016782

59. Hachlafi NE, Aanniz T, Menyiy NE, Baaboua AE, Omari NE, Balahbib A, Shariati MA, Zengin G, Fikri-Benbrahim K, Bouyahya A. *In vitro* and *in vivo* biological investigations of camphene and its mechanism insights: A review. Food Rev Int. 2023;39(4):1799-826.

https://doi.org/10.1080/87559129.2021.1936007

	Relative percentage (%) content of dominant compounds from the corresponding EOs										Reference	
Species	α-Pinene	Camphene	β -Pinene	δ-3-Carene	Limonene+β- Phellandrene	Bornyl acetate	(E)-Caryophyllene	α-Humulene	Germacrene D	Thunbergol		
P. halepensis	3.4	tr	0.3	1.1	0.4	0.1	32.2	5.9	1.0	29.2	[5]	
P. heldreichii	23.8	0.7	8.8	-	34.4	tr	11.1	1.1	17.3	-		
P. mugo	18.0	3.0	4.1	21.3	7.6	5.1	5.0	0.8	5.6	-		
P. nigra	35.4	0.9	10.0	0.1	2.8	0.6	7.4	1.3	28.1	-		
Р. реисе	49.3	8.1	13.0	0.1	5.5	7.7	1.8	0.2	6.5	-	[4]	
P. sylvestris	41.9	4.7	3.2	3.6	2.0	1.9	6.0	1.2	3.0	-		
A. alba	12.4	10.9	22.3	tr	17.0	8.8	5.7	2.6	0.9	-		
A. x borisii-regis	11.6	7.0	21.1	tr	12.1	5.7	8.0	4.5	2.0	-	[6]	
A. cephalonica	29.2	9.8	35.5	tr	12.4	2.4	2.0	0.8	-	-		

Supplementary Table S1. Ten dominant compounds contained in the EOs of Abies and Pinus species.

tr - trace<0.1%; not detected compounds are marked as (-); compounds that are >7% are in bold